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FastBlue™ Gel Staining Reagent

Cat no. PT-P575-500 ml

Description:

FastBlue™ Gel Staining Reagent is a convenient alternative to traditional Coomassie Blue staining procedures, based on a colloidal G250 formulation. Environmentally friendly, this ready-to-use stain does not contain methanol, acetic acid and TCA and does not require hazardous solvents for destaining. The protein bands are visible directly during the staining process in 3 min.

Generally the gel can be documented or stored directly without destaining. If optimal sensitivity is required, a simple and quick destaining with water yields clear background. The sensitive of detection is up to 8 ng under standard procedure.

Instructions:

Pre-wash: place the SDS-PAGE gel in a clean tray and rinse 5 min x 3 with gentle shaking, use 300-400 ml ultra-pure water per 8x 10 cm mini gel.

Note*: 1. SDS will inhibit the binding of dye with protein, it is very important to use large amount of water to remove SDS.*

2. If using 200 ml water to wash, 10 min x 3 wash will be recommended for maximum sensitivity.

3. A simple 5 min pre-wash is usually sufficient for native gel.

4. 10 min x 3 wash will be recommended for > 1mm thicker gel or > 15% gel.

5. For larger gel, use about 5 ml water per cm^2 gel to wash.

Stain: *Mix the FastBlue stain by inverting the bottle several times before use.*

Use 12-20 ml (depending on tray size, use stain solution just cover the gel) FastBlue Stain for an 8x 10 cm mini gel with gentle shaking for 30 min to 1 hr. The signals can be seen directly in the tray in 3 min.

Note*: 1. You can leave the gel overnight when necessary, which will not affect the sensitivity and background.*

2. Usually the background is very low in the staining solution, gel can be documented, stored or dried directly without destaining.

3. If destaining step is not necessary, do not rinse with water or the background will turn a little blue when rinsing with water.

Optional: Destain-background clearance

Discard the staining reagent and wash with water by alternately following steps:

A. Fast step by microwave or pre-warmed 50-60°C water

1. Microwave: add 100 ml ultra-pure water per mini gel in the *microwavable stray*, microwave 30 s then gentle shaking for 5 min. Repeat one or two more times to get a clearest background.

2. Pre-Warmed water: add 100 ml pre-warmed 50-60°C water to the gel in the tray, gentle shaking for 5 min. Repeat one or two more times to get a clearest background.
   Note: For larger gel, use 1.5 ml water per cm^2 gel to destain, microwave the water until water temperature is around 60°C; or using pre-warmed water to destain.

B. Slow step with water at room temperature
   Add 200 ml ultra-pure water to the gel in the tray, gentle shaking for 1-2 hour. Change water for 2 or 3 times during incubation is recommended for clearer background.
   Note: For larger gel, use 1.5 ml water per cm^2 gel to destain.

Storage of the gel:

1. The gel without destaining can be stored in original staining solution in plastic zip bag at 4°C for several weeks, do not rinse with water which will change pH and produce background.
2. The gel after destaining can be stored in the water at 4°C for several weeks. You may insert the gel into plastic zip bag with ultra-pure water and store at 4°C. Do not store the gel at RT for more than two days.

Destaining Protein Bands for MS Analysis:

General guidelines are provided below for destaining the protein bands prior to MS analysis. Contact your MS facility or the protein core facility for detailed protocols.

1. Excise the protein band of interest from the gel using a clean scalpel and destain with 10-30% ethanol or 20-30% acetonitrile for 10-15 minutes or until clear.
2. Rinse the gel piece in ultrapure water and proceed for MS analysis.

For Research Using Only.

Please do not hesitate to contact us if you have any questions.

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